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Estimation of α and β Peptide Bonds in Thermal **Poly(aspartic acid) by Potentiometric Titration**

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Summary

A petentiometric titration method for the esti $mation$ of the configuration (α and β peptide bonds, I and II) of poly(aspartic acid) prepared by thermal polycondensation is presented. Experimental results of the potentiometric titration of various samples differing in the molar fraction of I and II are confronted with the results of ¹³C NMR spectroscopy. Both methods lead to comparable results.

Introduction

Poly(Aspartic acid), poly(Asp), prepared by thermal polycondensation, and its derivatives are being studied as potential drug carriers, plasma expanders, models in the investigation of the fate of polymers in the body (KALAL et al. 1978) and in some other model studies (cf. PIVCOVA et al. 1981). Poly(Asp) is also regarded by some authors as a kind of a primitive prebiotic protein (proteinoid, FOX and DOSE 1977). It has been established that aspartic acid is linked in the main chain by the α (I) and β (II) peptide bonds:

> NH−CH−CO ~ NH−CH−CH₂−CO ~
│ │ │ │ │ │ │ ○
│ │ │ │ │ ○ СООН COOH

> > I II

The mole ratio of I and II may be determined by 13 C NMR (PIVCOVA et al. 1981) and according to some preliminary results also by IH NMR spectroscopy (MATSUYAMA et al. 1980). An alternative method is potentiometric titration (henceforth PT) which is used by Harada and coworkers. Analysing this method (SAUDEK 1981), we have shown that its treatment of the experimental data was not correct. Erroneous values of pK of I and II and of their molar fraction were obtained by extrapolation of the PT data and by the evaluation of the inflection

point in the PT curve (KOKUFUTA et al. 1977). Con sequently, the calculation of the molar fraction of II based on these pK values (HARADA et al. 1978, MATSUYAMA et al. 1980) also provided incorrect results. We propose, in the present communication, another approach.

Experimental

The samples of poly(D,L-aspartic acid), poly(Asp), were prepared by hydrolysis of poly(D,L-succinimide) (product of the thermal *polycondensation* of aspartic acid) under various pH. Their preparation and characterisation will be described in detail elsewhere (PIVCOVA et al.). The molar fraction of II and intrinsic viscosity are given in Table I.

PT was carried out with a Mettler automatic titrator equipped with the Radiometer combined electrode GK2402C under nitrogen atmosphere at 25°C. pH scale was calibrated with Radiometer buffers pH 7.00, 4.01 and 2.68. About 20 mg of a polymer was dissolved in 50 ml of 2 molal NaCI, 2.5 ml of 0.1M NaOH was added and the solution was titrated with 0.1M HCI. The titrant was added in such portions that the change of pH was 0.04-0.1 unit, each portion 10 s after dpH/dt < 5xi0 -4 pH unit/s. The exact concentration of the polymers was determined by PT and elemental analysis (N). The activity coefficient of H^+ ion in 2 molal NaCI was determined by titration of the solvent.

Data Treatment

An equation describing the dependence of pH on the degree of ionization (a) upon neglection of any polymer effect has been derived (SAUDEK, 1981):

$$
\alpha = \frac{K_1 k}{K_1 + H} + \frac{K_2 (1 - k)}{K_2 + H}
$$
 (1)

$$
\alpha = \frac{[Na^+] - [OH^-] + H}{C}
$$
 (2)

where $[Na^+]$, $[OH^+]$, H, C, k, K₁, K₂ are respectively the concentrations of added NaOH, hydroxyl ions, hydrogen ions and aspartic acid units, molar fraction of II and dissociation constants (defined by concentrations) of carboxyls of I and II. The polymer effect may be reduced for $\alpha < 0.7$ (MORAWETZ 1965, SAUDEK 1981) by the presence of a neutral salt, in this work with 2 molal NaCI. A set of n experimental points is obtained in the equilibrium PT: H_i , calculated from the measured pH and the activity coefficient of H⁺, and α_i , calculated according to Eq. (2) from the added

titrant volume, pH and activity coefficient of H^+ , $i=1,2,\ldots,n$. K₁, K₂ and k may be evaluated by the least squares method: A set of n equations (4) is obtained after substitution (3) and introduction of the experimental data pairs α_i and H_i in Eq. (1):

$$
x = K_1 K_2 , \qquad y = K_1 + K_2 , \qquad z = K_2 k - K_1 k - K_2
$$
 (3)

$$
x (\alpha_{i}^{-1}) + y \alpha_{i} H_{i} + z H_{i} + \alpha_{i} H_{i}^{2} = \varepsilon_{i}
$$
 (4)

where ${\tt E_i}$ is residuum of the determination. The minimalization condition for the residual sum of squares $\sum_{i=1}$ ε_i^2 leeds to following equations: n _n n n n $x \geq (\alpha_i - 1) + y \geq (\alpha_i - 1) \ln_i \alpha_i + z \geq \ln_i (\alpha_i - 1) + \sum_{i} \alpha_i \ln_i (\alpha_i - 1) = 0$ i=1 i=1 i=1 i=1 i=1 $\sum_{i=1}^{L} (\alpha_i - 1) H_i \alpha_i + \sum_{i=1}^{L} H_i^2 \alpha_i^2 + \sum_{i=1}^{L} H_i^2 \alpha_i + \sum_{i=1}^{L} \alpha_i^2 H_i^3 = 0$ (5) i=I $n = \frac{n}{2}$ $\frac{n}{2}$ $\frac{n}{2}$ $\frac{n}{2}$ $\frac{n}{2}$ $\frac{n}{2}$ $\frac{n}{2}$ $\alpha_{i=1}^{x}$ $\alpha_{i}^{H_{i}}$ $\alpha_{i+1}^{H_{i}}$ $\alpha_{i}^{H_{i}}$ $\alpha_{i}^{H_{i}}$ $\alpha_{i+1}^{H_{i}}$ $\alpha_{i+1}^{H_{i}}$ $\alpha_{i+1}^{H_{i}}$ $\alpha_{i+1}^{H_{i}}$

The solution of (5) provides x, y and z and, according to (4) , K_1 , K_2 and k.

The results may be further improved supposing that the set of m various poly(Asp) samples differs in k only, K_1 , K_2 being identical for all polymers. The least squares method leads to (m+2) equations (7) after the substitution (6):

$$
x = K_1 K_2 , \qquad y = K_1 + K_2 , \qquad z_h = K_2 k_h - K_1 k_h + K_2
$$
 (6)

$$
\sum_{h=1}^{m} \sum_{i=1}^{nh} (\alpha_{ih}^{-1})^2 + \sum_{h=1}^{m} \sum_{i=1}^{nh} \alpha_{ih} H_{ih} (\alpha_{ih}^{-1}) +
$$

$$
+\sum_{h=1}^{m} z_{h} \sum_{i=1}^{n_h} H_{ih} (\alpha_{ih} - 1) + \sum_{h=1}^{m} \sum_{i=1}^{n_h} \alpha_{ih} H_{ih}^2 (\alpha_{ih} - 1) = 0
$$

$$
x \sum_{h=1}^{m} \sum_{i=1}^{n_h} (\alpha_{ih}^{-1}) H_{ih} \alpha_{ih} + y \sum_{h=1}^{m} \sum_{i=1}^{n_h} \alpha_{ih}^{2} H_{ih}^{2} +
$$

$$
+\sum_{h=1}^{m} z_{h} \sum_{i=1}^{nh} \alpha_{ih} H_{ih}^{2} + \sum_{h=1}^{m} \sum_{i=1}^{nh} \alpha_{ih}^{2} H_{ih}^{3} = 0
$$
 (7)

$$
x \sum_{i=1}^{n_h} (\alpha_{ih} - 1) H_{ih} + y \sum_{i=1}^{n_h} \alpha_{ih} H_{ih}^2 + z \sum_{i=1}^{n_h} H_{ih}^2 + \sum_{i=1}^{n_h} \alpha_{ih} H_{ih}^3 = 0
$$

where h=1,2,...,m, i=1,2,...,n_h, $k_{\sf h}$ is the molar fraction Qf II in h-th poly(Asp) sample, $\alpha_{\rm ih}$, H_{ih} and $n_{\rm h}$ are respectively the i-th degree of ionization, i-th hydrogen ion concentration and number of the experimental points obtained by PT of h-th sample.

The calculations were performed on a Wang 2200 calculator.

Results and Discussion

The calculation was carried out for α_1 in the interval 0.08-0.7; smaller α_i were not used because of a relatively great experimental error (the accuracy of pH determination at low values is low and small error in pH causes great error in α_i at low α_i); higher α_i were not used because the polymer effect may not be compensated enough for these values even in a 2 molal NaCI. About 40 experimental points were used from one PT experiment.

Table I presents the results obtained according to Eqs (5) (method A - each PT evaluated separately) and (7) (method $B - PT$ of all 5 samples evaluated together). The correlation coefficient of experimental α_i and regression curve was greater than 0.99 standard deviation for $\alpha_{\rm i}$ lower than 10 $^{-5}$ in all cases. The supposition that K₁ and K₂ do not differ for various poly(Asp) samples of different k and molecular weight is acceptable. The results of the method B are based on a higher number of experimental points and may be regarded as more pertinent. Figure la shows the titration curve of poly(Asp)III, Fig. Ib represents its modified titration curve. The regression curve fits the experimental points very good. The deviations are somewhat more apparent from the plot usually employed for the PT curves of polymers (Fig.lb). Even the points measured for α_i <0.08, not used in the evaluation, do not substantially deviate from the regression curve. The figures also indicate that the polymer effect was compensated enough for $\alpha_i < 0.7$ -0.8. Similar agreement was obtained with all the other samples. Equation (I) is a good representation of the PT behaviour of poly (Asp) for α_i <0.7-0.8.

Table I presents for comparison the results obtained by means of NMR spectroscopy. The k values determined by PT are somewhat smaller but lie within

476

477

Fig.1. (a) Titration curve of poly(Asp)III. (b) Modified plot pK_{app} = pH+(1- α)/ α vs. α of the titration curve of poly(Asp)III. The dots represent experimental points, the line the regression curve.

the range of the experimental error, in good agreement with those obtained by NMR spectroscopy. PT is of course not the best method for the problem studied. Evidence of I and of II was given only indirectly by PT and assuming that the polymer effect was compensated sufficiently. NMR spectroscopy provides evidence of I and II directly. Nevertheless, the comparison in Table I shows that PT leads to reliable results.

TABLE I

Analysis of $poly(Asp)$ samples $(k - molar fraction of$ II, K₁, K₂ - dissociation constants of I and II, pr_1 = -log K₁, pr_2 = -log K₂)

a Preparation and characterization: PIVCOVÁet al. b^{F1} Eparation and shares of I and I pH = 7.3 CEvaluated from the relative intensities of the ¹³C NMR CH₂ bands at pH = 9.2 (PIVCOVA et al. 1981) dEvaluated according to Eqs (5)

eEvaluated according to Eqs (7)

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478